

Premix Ex Taq™ DNA Polymerase Hot Start (Cat.# RR030A)

## Application: Amplification of GC-rich DNA Fragments using TaKaRa LA Taq® or Ex Taq® DNA Polymerase

When amplifying template DNA that is GC-rich, conventional Taq DNA Polymerase often does not perform well. The superior amplification efficiency of TaKaRa LA Taq® and TaKaRa Ex Taq® allows amplification of difficult DNA targets, such as those with high GC content.

This application example illustrates the ability of these enzymes to amplify a 3 kb fragment that has 70% GC content.

### Methods:

**Template:** VIII-5-pUC118 (a ~3 kb GC-rich fragment inserted into the Hind III site of pUC118)

**Amplification size:** ~3 kb

**Primers:** M3-30 (an extended M3 Primer)  
5'-CAGTCACGACGTTGTAACGACGGCCAGT-3'  
RV-32 (an extended RV Primer)  
5'-GATAACAATTTACACAGGAAACAGCTATGAC-3'  
(10 pmol of each primer was used per 50 µl PCR reaction)

**dNTP mixture:** 7-deaza-dGTP was used instead of dGTP

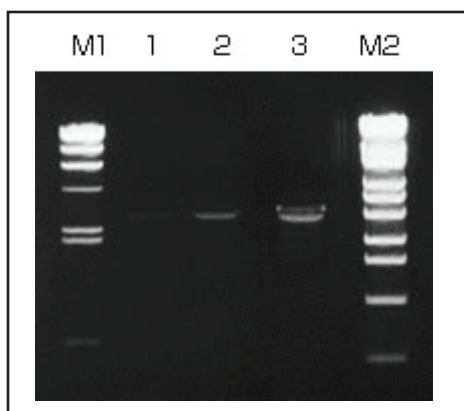
### PCR conditions:

94°C, 1 min.  
↓  
98°C, 20 sec. } 30 cycles  
68°C, 5 min. }

**Thermal cycler:** TaKaRa PCR Thermal Cycler 480\*

**Enzymes:** TaKaRa LA Taq® – LA PCR Kit Ver. 2.1 (Cat.# RR013A)  
TaKaRa Ex Taq® – Premix Ex Taq™ DNA Polymerase Hot Start Version (Cat.# RR030A)

### Results:



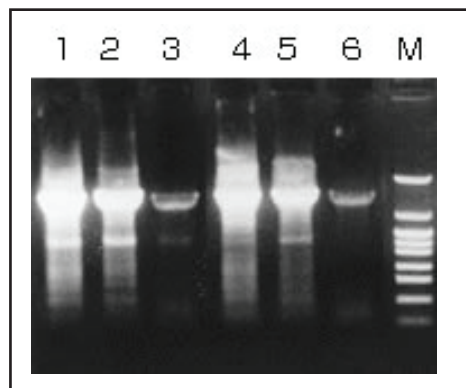
**Figure 1.** Amplification of a GC-rich fragment using various DNA polymerases. PCR products were amplified using either conventional Taq (lane 1), Ex Taq® (lane 2), or LA Taq® (lane 3) and were run on a 1% L03 agarose gel (8 µl PCR reaction per lane). Lanes M1 and M2 contain molecular weight markers (λ-Hind III digest and λ-EcoT14 digest, respectively).

\*Not available in all geographic locations. Check for availability in your region.

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## Application: GC-rich DNA Fragments using *TaKaRa LA Taq*<sup>®</sup> or *Ex Taq*<sup>®</sup> DNA Polymerase (page 2)

Using 7-deaza-dGTP instead of dGTP in the PCR reaction can improve amplification of GC-rich templates. In addition, further optimization using a mixture of dGTP and 7-deaza-dGTP (or dITP) can also improve amplification efficiency (Figure 2).



**Figure 2.** Amplifications of a GC-rich fragment using various ratios of dGTP and 7-deaza-dGTP or dITP. PCR products were amplified using either *LA Taq*<sup>®</sup> (lanes 1–3) or *Ex Taq*<sup>®</sup> (lanes 4–6) and were run on a 1% L03 agarose gel (10  $\mu$ l). The PCR reactions contained various combinations of dGTP, dITP, and 7-deaza-dGTP as indicated in the table above. Lane M contains a  $\phi$ Y molecular weight marker.

Lane 1: dGTP:dITP = 3:1

Lane 2: dGTP 7-deaza-dGTP = 3:1

Lane 3: 7-deaza-dGTP alone

Lane 4: dGTP:dITP = 3:1

Lane 5: dGTP:7-deaza-dGTP = 3:1

Lane 6: 7-deaza-dGTP alone

### Conclusions:

A GC-rich fragment that was not efficiently amplified using conventional *Taq* could be successfully amplified with both *TaKaRa Ex Taq*<sup>®</sup> and *TaKaRa LA Taq*<sup>®</sup> DNA polymerases.

Amplification of GC-rich target sequences can be improved by replacing dGTP with dITP or 7-deaza-dGTP. Depending on fragment to be amplified, the optimal ratio of dGTP to 7-deaza-dGTP (or dITP) may vary. Therefore, we recommend evaluating various ratios to determine the ideal conditions.